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1 **Surface-modified phillipsite-rich tuff from the Campania region (southern Italy)**
2 **as a promising drug carrier: An Ibuprofen sodium salt trial**

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19 **ABSTRACT**

20 The encapsulation and delivery of drugs often involves the use of expensive
21 microporous materials, and we have investigated the potential for natural zeolites from
22 the widespread volcanic formations of southern Italy as alternatives to these carriers.

23 Surface-modified natural zeolites (SMNZs) with diverse micellar structures (patchy

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24 and complete bilayers) were obtained by using different cationic surfactants
25 (cetylpyridinium chloride (CP-Cl), benzalkonium chloride (BC-Cl),
26 hexadecyltrimethylammonium chloride (HDTMA-Cl) and bromide (HDTMA-Br) with
27 phillipsite-rich tuff from the Campania region (southern Italy). Loading and release
28 kinetics tests of sodium Ibuprofen (IBU) were carried out with organo-phillipsite
29 composites using Fourier-transform infrared spectroscopy (FTIR) and thermal analysis
30 coupled with evolved gas analysis (EGA). Results from these tests were
31 mathematically modeled to evaluate IBU adsorption and release mechanisms.
32 The maximum loaded amount of IBU was attained for organo-phillipsite modified with
33 HDTMA-Br (PHB), which showed a complete bilayer micellar structure. Whenever a
34 patchy bilayer micellar structure formed, the lowest adsorptions of IBU were observed.
35 Equilibrium adsorption results were fit using Langmuir, Sips and Toth models. Pseudo
36 first-order and pseudo second-order fits to the loading kinetic data provided significant
37 Goodness of Fit. Good fits to the release kinetic data were obtained using first-order
38 and Weibull equations, shedding new light on the release mechanism of IBU from
39 phillipsite. The active amount of IBU on the modified zeolite surface was almost
40 totally available for pharmaceutical purposes.

41

42 **Keywords:** phillipsite; Neapolitan Yellow Tuff; Ibuprofen sodium salt;
43 functionalization; SMNZ; carrier; drug delivery; SIPS model; TOTH model.

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INTRODUCTION

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46 Recent research has demonstrated that natural and surface-modified zeolites containing
47 cationic surfactants are selective towards some pharmacological molecules (i.e.,
48 diclofenac sodium, diclofenac diethylamine, and Ibuprofen). Available data suggest
49 that zeolites represent a low cost and easily accessible natural excipient that can be
50 used in biomedicine as carriers for drug delivery (Cerri et al. 2004, 2016, Krajišnik et
51 al. 2013b, 2015, 2010a, 2010b, 2011, 2013a; Cappelletti et al. 2017; de Gennaro et al.
52 2015, 2016; Jančićjević et al. 2015; Marković et al. 2016, 2017; Pasquino et al. 2016;
53 Serri et al. 2016, 2017). Based on the premise that such natural carriers are non-toxic
54 (Mercurio et al. 2012, 2016b; Krajišnik et al. 2013b; Cerri et al. 2016), several aspects
55 of their performance should be clarified before modified zeolites can be used on an
56 industrial scale. In order to evaluate their functional therapeutic performance, it is
57 particularly important to expand the data for a breadth of different systems such as
58 zeolite vs. surfactant vs. active substance. Careful evaluation of loading and release
59 performances under controlled conditions is also required (Krajišnik et al. 2015, 2010a,
60 2010b, 2011, 2013a, 2013b, de Gennaro et al. 2015, 2016, 2017; Jančićjević et al.
61 2015; Marković et al. 2016, 2017; Pasquino et al. 2016; Serri et al. 2016, 2017; Izzo et
62 al. 2017). Several recent studies have documented the efficacy of zeolites as
63 pharmacological carriers. Serri et al. (2017) demonstrated that granulate could be
64 formed using a functionalized natural clinoptilolite, suitable for the release of sodium
65 diclofenac. Krajišnik et al. (2016) demonstrated that non-steroidal anti-inflammatory
66 drugs (NSAID), for example IBU, benefit particularly from use with therapeutic
67 devices providing prolonged release, such as clinoptilolite-rich carriers. For loading
68 purposes, Krajišnik et al. (2016) used clinoptilolite modified with two surfactants

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69 (cetylpyridinium chloride and benzalkonium chloride) at different concentrations, and
70 their results showed that different adsorbed amounts of IBU depended on the type
71 and/or the amount of surfactant used to functionalize the zeolite-rich carrier. In *vitro*
72 release tests also demonstrated that IBU was released from the composite material for
73 up to eight hours, as illustrated by mathematical models Bhaskar (Bhaskar et al. 1986)
74 and Higuchi (Higuchi 1963). Krajišnik et al. (2016) hypothesized that the adsorption
75 process involved hydrophobic-hydrophilic interactions between the drug molecules
76 and the surfactants on the zeolite surface.

77 In order to meet the demand of the pharmaceutical industry searching for new and
78 inexpensive carriers for the encapsulation and delivery of drugs (Tan et al. 2013), we
79 investigated natural and widespread phillipsite-rich rock from southern Italy
80 (Campania region) as alternatives to the usual expensive prepared mesoporous
81 materials. This geomaterial, widely occurring in the Neapolitan Yellow Tuff (NYT)
82 Formation and the Campanian Ignimbrite (CI) Formation, has been studied in detail
83 (de Gennaro et al. 1992, 1995, 1999, 2000a; Colella et al. 1998, 2013, 2017; Langella
84 et al. 2002, 2013; Cappelletti et al. 2003; Calcaterra et al. 2004; Buondonno et al.
85 2008; Morra et al. 2010), and existing data support applications in the areas of cation
86 exchange (Pansini et al. 1996; Colella et al. 1998; Gatta et al. 2015), animal feeding
87 (Mercurio et al. 2012, 2016b), oenotechnical (Mercurio et al. 2010, 2014, 2016a), soil
88 remediation (Coppola et al. 2003), production of lightweight aggregates (de Gennaro et
89 al. 2004, 2005, 2007, 2008; Dondi et al. 2016), and ceramic manufacturing (de
90 Gennaro et al. 2003; Cappelletti et al. 2011).

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91 Application of natural zeolites in the pharmaceutical field requires evaluation of the
92 performance of functionalized zeolites as carriers for drug delivery. Modification of
93 the zeolite surface was performed in our studies using four different surfactants
94 (cetylpyridinium chloride, benzalkonium chloride, hexadecyltrimethylammonium
95 chloride and bromide) at a concentration equal to 200% of the external cation exchange
96 capacity (ECEC) of the zeolite. The amount of surfactant was chosen based on
97 published results (Li and Bowman 1997; de Gennaro et al. 2016; Cappelletti et al.
98 2017) that demonstrated that this concentration guarantees the almost complete
99 formation of a bilayer of the surfactants on the zeolite surface.
100 In order to clarify the properties of this specific phillipsite-IBU composite material,
101 equilibrium isotherms as well as IBU loading and release kinetics were measured.
102 Basic characterization of the composite material was obtained using thermal analysis
103 (STA coupled with EGA) and FT-IR spectroscopy.

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MATERIALS

106 A phillipsite-rich tuff from the Campania Region (southern Italy) and four cationic
107 surfactants were selected for the preparation of surfactant modified natural zeolites
108 (SMNZs) and were used for subsequent IBU sodium loading and release tests.
109 Mineralogical and technological characterization of the natural zeolite-rich material
110 has been reported by Cappelletti et al. (2017) and is only briefly summarized here.
111 NYT phillipsite-rich rocks from Marano (Savanelli quarry, Naples, Italy; hereafter
112 PHI_SAV) contain about 70 wt.% phillipsite with minor chabazite (~5 wt.%) and
113 analcime (~3 wt.%) (de' Gennaro et al. 2000b). This phillipsite from the PHI_SAV

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114 sample has a low Si/Al ratio (approximately 2.5) and it is characterized by a high K^+
115 content, which reflects the typical composition of trachytic rocks from Campi Flegrei
116 (southern Italy) (Morra et al. 2010). It contains subordinate amounts of Ca^{2+} and Na^+ .
117 The experimental ECEC value (0.144 mEq/g) was evaluated using the method
118 suggested by de Gennaro et al.(2014), which involves contacting a zeolite sample (2.5
119 g) with 10 mL of 20-mM surfactant solution in a 50-mL polyallomer centrifuge tube,
120 shaking at 100 rpm for 24 h at 25 °C. The sample was then centrifuged and the
121 supernatant analyzed for the anion concentration by HPLC.
122 Leaching tests confirmed that the available contents of heavy metals such as Ni, As,
123 Cd and Pb were all below 20 mg/kg, confirming the correlation with the total amount
124 of these elements in the whole rock (Mercurio et al. 2012).
125 PHI_SAV was surface modified using cetylpyridinium chloride (CP-Cl),
126 benzalkonium chloride (BC-Cl), hexadecyltrimethylammonium chloride (HDTMA-Cl)
127 and bromide (HDTMA-Br). The resulting SMNZs were labeled as PCC, PBC, PHC
128 and PHB, respectively. Anion exchange capacity (AEC) values for these were ~0.09
129 mEq/g for PBC and PC, and ~0.110 mEq/g and 0.140 mEq/g for PHC and PHB. These
130 results are consistent with the different micellar structure (patchy and complete bilayer)
131 obtained using different surfactant molecules (Cappelletti et al., 2017).
132 These organo-phillipsite composites were used for loading and release tests of sodium
133 IBU. Chemical characteristics of surfactants and IBU are presented in Figures 1 and 2.

134

135

METHODS

136 **SMNZs preparation**

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137 In order to obtain SMNZs for the IBU loading tests, a suspension containing PHI_SAV
138 and each of the selected surfactants, having an initial concentration equivalent to 200%
139 of the ECEC, was prepared. The suspension was mixed at 3,000 rpm using a FALC
140 AT-M 20 stirrer at ~ 50 °C for ~ six hours, with a solid to liquid (S/L) ratio of 1/40.
141 The suspensions were then filtered and the obtained SMNZs were washed with
142 ultrapure water to remove the unnecessary surfactant and were then dried at room
143 temperature for 24 hours.

144

145 **Drug loading tests**

146 Maximum IBU adsorption capacities of SMNZs were determined by means of
147 equilibrium and kinetics adsorption tests. The pH of each suspension was adjusted to
148 7.4 with a sodium tetraborate buffer solution (CAS No. [1303-96-4]) as this pH value
149 allows the IBU sodic salt to remain in its ionic form (Krajišnik et al. 2015; Oh et al.
150 2016).

151 For the equilibrium adsorption measurements, 100 mg of each SMNZ were treated
152 with 20 mL of IBU solution, having concentrations ranging between 50 and 1000 mg/L,
153 and continuously stirred in 25 mL Nalgene centrifuge tubes at room temperature. After
154 four hours (sufficient time to attain equilibrium based on the kinetic tests), samples
155 were centrifuged to separate solid from liquid and the supernatant was analyzed using
156 an AquaMate UV-VIS spectrophotometer. The IBU calibration curve ($R^2=0.999$) was
157 obtained using the adsorption band at 224 nm, in accordance with (Sena et al. 2007;
158 Gondalia et al. 2010; Joshi et al. 2011). Each measurement was made in triplicate.

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159 Kinetic studies were carried out in batch experiments at room temperature and under
160 continuous stirring, by shaking 100 mg of each sample with 20 mL of IBU solution
161 (400 mg/L). To avoid variation of this solution/SMNZ ratio the mixture was placed
162 into 25 mL high density polyethylene centrifuge tubes. A test tube was withdrawn
163 (after 5', 10', 15', 30', 60', 90' ...240), centrifuged, and the non-adsorbed amount of
164 IBU was determined in the supernatant.

165

166 **Drug release tests**

167 IBU *in vitro* release experiments were carried out using IBU-loaded SMNZs dispersed
168 in simulated intestinal fluid (SIF), prepared as described in the United States
169 Pharmacopoeia 25 Ed. and later editions (USP-NF 2002, 2003). 15 mg of IBU-loaded
170 SMNZs were added to 10 mL of SIF and placed in a thermostatic bath at 37 °C under
171 continuous stirring (100 rpm). Samples were kept in the thermostatic bath until the
172 released amount of the drug reached a plateau. The IBU-loaded SMNZ/SIF ratio was
173 chosen in order to ensure sink conditions. 5 mL aliquots of supernatant were
174 withdrawn at fixed interval times (30 min), centrifuged at 9,000 rpm (room
175 temperature, 5 min), and replaced by the same volume of fresh medium. IBU was
176 quantified by spectrophotometric assay (Aquamate UNICAM) at 224 nm. Experiments
177 were run in triplicate.

178

179 **Mathematical modeling**

180 To evaluate IBU adsorption and release mechanisms of SMNZs, adsorption isotherms
181 and kinetic measurements were fit using several mathematical models proposed in the

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182 literature (Hinz 2001; Ho 2004, 2006; Ho et al. 2005; Limousin et al. 2007; Dash et al.
183 2010; Foo and Hameed 2010; Wang and Peng 2010; Chen 2013; Yadav et al. 2013).
184 Fit parameters were determined by non-linear regression using the Generalized
185 Reduced Gradient algorithm (Gabriele and Ragsdell 1977), based on the assumption
186 that non-linear regression provides the best fit results (Ho et al. 2005; Lin and Wang
187 2009; Chen 2013; Markandeya and Kisku 2015).
188 Model fit and applicability were evaluated by considering determination coefficients
189 (R^2), as well as other statistical methods for non-linear regression (Costa et al. 2003;
190 Spiess and Neumeyer 2010) such as the Akaike Information Criterion (AIC) (1) and
191 the Bayesian Information Criterion (BIC) (2). These methods take into account the
192 number of parameters of the mathematical model and the number of experimental data
193 and can be expressed as follows:

194

$$195 \quad AIC = 2p - 2\ln(L) \quad ; \quad (1)$$

196

$$197 \quad BIC = p\ln(n) - 2\ln(L) \quad , \quad (2)$$

198 where p is the number of parameters and n is the sample size. $\ln(L)$, the maximum log-
199 likelihood of the estimated model (Spiess and Neumeyer 2010), was calculated as
200 follows: $\ln(L) = 0.5 * [-N * (\ln 2\pi + 1 - \ln N + \ln \sum_{i=1}^n x_i^2)]$, where x_i are the
201 residuals from the nonlinear least squares fit and $N =$ the number of residuals.

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203 **Thermal analyses**

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204 Thermal analysis (thermogravimetry and differential scanning calorimetry, TG/DSC)
205 was performed on phillipsite-rich material, cationic surfactants, IBU, SMNZs and IBU-
206 loaded SMNZs using a NETZSCH STA 449 F3 Jupiter instrument with alumina
207 crucibles. Samples were heated from room temperature to 1050 °C at a heating rate of
208 10 °C/min in an ultra-pure air atmosphere (N₂/O₂ = 80/20; flow 60 mL/min). EGA
209 were carried out by FTIR with a BRUKER Tensor 27 instrument, coupled to the STA
210 449 F3 instrument by a transfer line heated to 200 °C.
211 Netzsch Proteus 6.1.0 (NETZSCH-Gerätebau GmbH) and Opus 7.2 (Bruker Optics
212 GmbH) software packages were used for data analysis.

213

214 **FTIR**

215 The phillipsite-rich material, cationic surfactants, IBU, SMNZs and IBU-loaded
216 SMNZs were analyzed using a Bruker Alpha FTIR spectrometer in attenuated total
217 reflectance (ATR)-mode with 128 scans at a spectral resolution of 4 cm⁻¹ in the
218 spectral range of 400-4000 cm⁻¹. Spectra were analyzed with Opus 7.2 software
219 (Bruker Optics GmbH).

220

221 **RESULTS AND DISCUSSION**

222 **Characterization of SMNZs**

223 *FTIR spectroscopy*

224 The infrared spectrum of PHI_SAV shows the typical absorption bands of zeolite-rich
225 tuffs (Cappelletti et al. 2017), including absorption bands associated with internal and
226 external framework vibrations of primary building units (PBUs) and secondary

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227 building units (SBUs) of zeolites (Karge 2001; Byrappa and Kumar 2007; Mozgawa et
228 al. 2011). A strong band at $\sim 1000\text{ cm}^{-1}$ (Table 1 and Fig. 3) was assigned to
229 asymmetric stretching Si-O vibrations. Bands at ~ 780 and $\sim 719\text{ cm}^{-1}$ were assigned to
230 asymmetric and symmetric stretching T-O-T vibrations, respectively, and the
231 remaining bands are generally referred to bending vibrations characteristic for silicates,
232 also occurring in the sample, such as pyroxene (3.3 wt.%), K-feldspar (13.3 wt.%) and
233 traces of mica. The weak, broad band at $\sim 3419\text{ cm}^{-1}$ and the band at $\sim 1638\text{ cm}^{-1}$
234 suggest the presence of hydrogen-bonded H_2O molecules.
235 Moreover, a very weak broad band at $\sim 1456\text{ cm}^{-1}$ could indicate traces of carbonate
236 minerals (C-O asymmetric stretching in the carbonate ion).
237 IR vibrations associated with exchangeable cations usually occur in the far-IR region
238 ($200\text{-}50\text{ cm}^{-1}$) (Karge 2001), and any structural modification caused by external cation
239 substitution with cationic surfactants could not be detected by mid-IR spectroscopy.
240 On the other hand, FTIR spectra of SMNZs and IBU-loaded SMNZs can be clearly
241 distinguished from those of the starting materials by the presence of absorption bands
242 in the spectral range $3000\text{-}2800\text{ cm}^{-1}$ (Table 1; Fig. 3) (Barczyk et al. 2014), which
243 correspond to the strong bands (C-H stretching vibrations) observed both in surfactants
244 and IBU (Figures 1 and 2). These organic compounds also share two very weak bands
245 at $\sim 1487\text{ cm}^{-1}$ and $\sim 1468\text{ cm}^{-1}$ (C-H bending vibrations). However, IBU-loaded
246 SMNZs show increased intensity of C-H stretching vibrations (Krajišnik et al. 2010b,
247 2015), as well as the occurrence of additional bands at $\sim 1578\text{ cm}^{-1}$ and $\sim 1380\text{ cm}^{-1}$
248 (Table 1; Fig. 3) attributable to asymmetric and symmetric stretching vibrations in
249 carboxylate ions (Ambrogi et al. 2001; Wray et al. 2011).

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251 *STA coupled with FT-IR/EGA*

252 Thermal analysis has been widely used in the characterization of natural zeolites and
253 their modified forms (Sullivan et al. 1997; Bish and Carey 2001; Langella et al. 2003;
254 Krajišnik et al. 2011, 2013a, 2013b; de Gennaro et al. 2016; Marković et al. 2017). So-
255 called “zeolitic water” is found in the open cavities (channels and cages) of zeolites
256 (Coombs et al. 1997), where it can move more-or-less freely through the
257 interconnected cavities (Bish and Carey 2001; Langella et al. 2003). The phillipsite-
258 rich starting material shows the typical thermal behavior of a zeolite with a high kinetic
259 pore diameter (Bish and Carey 2001; Langella et al. 2003), as much of the weight loss
260 occurs below 250 °C with the maximum dehydration rate at ~ 176 °C (Fig. 4).
261 Dehydration continued gradually to ~ 550 °C, likely due to the presence of other
262 natural zeolites with lower kinetic diameters such as chabazite and minor analcime as
263 well as hydroxylated phases (micas). Cumulatively, the “zeolitic” H₂O evolving over
264 the 40-550 °C range is ~10 wt.%. This H₂O content is consistent with the high K⁺
265 content of the zeolites in the tuff. The presence of large monovalent cations, such as K⁺,
266 generally results in moderate or low H₂O contents (Bish 1984, 1988; Kranz et al. 1989;
267 Carey and Bish 1996; Fialips et al. 2005; Esposito et al. 2015).
268 H₂O evolved during dehydration was detected via FTIR/EGA (Fig. 4), along with a
269 weak emission of CO₂ attributable to the decomposition of calcite in the 550-800 °C
270 thermal range.

271 The thermal behavior of SMNZs and IBU-loaded SMNZs is mainly characterized by
272 the occurrence of two principal processes (Table 2). The first, occurring below 200 °C,

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273 is related to dehydration of the zeolitic component, and decomposition of cationic
274 surfactants was not significant in this temperature range (Fig. 4). The second process
275 (>200 °C) is linked to decomposition and subsequent combustion of surfactants
276 (SMNZs) and IBU (IBU-loaded SMNZs) (Table 2; Figs 1 and 2). Thus the total mass
277 loss for IBU-loaded SMNZs is greater than the mass loss for the starting material and
278 the SMNZs before drug adsorption (Table 2).

279 These results could attest the effective functionalization of zeolite and subsequent drug
280 loading although, it requires a further validation by technological performance.

281

282 **Technological performance of the phillipsite-IBU composite material**

283 *Equilibrium isotherms*

284 Table 3 and Figure 5 report the results of equilibrium adsorption measurements fit
285 using three mathematical models, Langmuir (Fig. 5a) (Langmuir 1916), Sips (also
286 called Langmuir-Freundlich, Fig. 5b), and Toth (Fig. 5c) equations.

287 The shape of the curves shows that the plateau, i.e., the maximum drug adsorption, was
288 reached for the initial IBU concentration of 400 mg/L, regardless of the type of
289 surfactant. Moreover, the maximum loaded amount was attained for the PHB (~29
290 mg/g), in agreement with complete bilayer formation, whereas lower adsorption was
291 achieved for PBC and PCC (21.2 mg/g) where the two surfactants formed only a
292 patchy bilayer at the zeolitic surface. The latter are in agreement with Cappelletti et al.
293 (2017), who emphasized the effects of the counterions on micellar structure formation.
294 The same behavior was also recorded for PHC, although it achieved a higher IBU
295 loaded amount. This may indicate that, for its benzene ring-free structure (different

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296 from benzalkonium and cetylpyridinium), the HDTMA-Cl can form a more
297 homogeneous patchy bilayer than the other two chlorinated surfactants, as found in
298 previous research (Cappelletti et al., 2017).
299 The Langmuir adsorption isotherm model is commonly used to fit the performance of
300 different bio-mineral adsorbents such as SMNZs (Li et al. 1998, 1999; Krajišnik et al.
301 2010a; Wang and Peng 2010; Cappelletti et al. 2017; de Gennaro et al. 2016; Marković
302 et al. 2017) and other geomaterials (Campbell and Davies 1995; Li and Bowman 1997,
303 1998, 2001; Janićijević et al. 2015), although it was originally developed to describe
304 gas-solid-phase adsorption onto activated carbon (Langmuir 1916; Foo and Hameed
305 2010). On the other hand, the Sips (1948) and Toth (1971) models were developed to
306 improve fits based on Langmuir isotherms (Hinz 2001; Limousin et al. 2007; Foo and
307 Hameed 2010).
308 The Langmuir (equation 3), Sips (equation 4), and Toth (equation 5) equations can be
309 expressed as follows:

310

$$311 \quad S = S_m \frac{KC_e}{1+KC_e}, \quad (3)$$

312

$$313 \quad S = S_m \frac{(KC_e)^n}{1+(KC_e)^n}, \text{ and} \quad (4)$$

314

$$315 \quad S = S_m \frac{KC_e}{[1+(KC_e)^n]^{\frac{1}{n}}}, \quad (5)$$

316 where S is the amount of solute adsorbed by the solid at equilibrium conditions (mg/g);
317 C_e is the concentration of solute in solution after equilibrium (mg/L); S_m is the

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318 maximum adsorption capacity at equilibrium (mg/g), K (L/mg) is the adsorption
319 intensity, a constant related to binding energy, and n is a fitting parameter related to
320 heterogeneity of the system. If $n = 1$, the Sips and Toth equations become a simple
321 Langmuir equation.

322 The Sips and Toth models generally provided improved Goodness-Of-Fit (GOF)
323 values observed as a reduction of AIC and BIC values, as well as an increase in the
324 determination coefficient R^2 (Table 3). Maximum IBU adsorption capacities (S_m) of
325 22.0 mg/g, 20.4 mg/g, and 20.4 mg/g was calculated for sample PCC according to the
326 Langmuir, Sips, and Toth isotherm equations, respectively (Table 3). Both the Sips
327 (1948) and Toth (1971) provided best fits ($R^2 = 0.987$; AIC = 23.1; BIC = 22.9) for
328 PCC, but no significant differences were found between these three models for IBU
329 adsorption by PBC (Table 3).

330 Greater maximum adsorption capacities were obtained for PHC and PHB compared
331 with PCC and PBC, and the Toth equation provided the best fit to the data (Table 3 and
332 Fig. 5), particularly for PHB.

333 The Toth fit parameters yield an amount of adsorbed IBU of 24.4 mg/g for PHC and
334 28.8 mg/g for PHB. The better performance of PHB can be explained by the tendency
335 of this composite to form a more compact bilayer sheet micelle due to the presence of a
336 Br counterion (Li and Bowman 1997; Cappelletti et al. 2017; de Gennaro et al. 2016).

337 The general improvement of the GOF with the Sips and Toth equations instead of the
338 Langmuir can be explained by taking into account the theoretical basis of the Langmuir
339 isotherm model (Langmuir 1916; Limousin et al. 2007; Foo and Hameed 2010). The
340 Langmuir model is strictly intended for materials where sorption mechanisms are fully

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341 associated with adsorption processes. The model is therefore inappropriate for SMNZs
342 where sorption of NSAIDs is controlled by multiple mechanisms, including both
343 external anionic exchange and partition into the hydrophobic portion of the micelle
344 (Krajišnik et al. 2010a, 2010b, 2011, 2013a, 2013b, 2015; de Gennaro et al. 2015;
345 Marković et al. 2016; Pasquino et al. 2016; Serri et al. 2017). In case of multiple
346 sorption mechanism, although the adsorption mechanism always predominates (de
347 Gennaro et al. 2015; Krajišnik et al. 2015), partition may influence the sorption of IBU
348 by SMNZs, especially when IBU molecules can change their ionic form (e.g. by pH
349 variations). Thus, from a mathematical point of view, the Sips or Toth equations fit the
350 adsorption data better than the Langmuir model and they provide a well-defined
351 equilibrium asymptotic plateau, useful for the identification of the real maximum
352 adsorption capacity of SMNZs. In this way, equilibrium isotherm modeling can be
353 used to optimize the appropriate initial concentration of IBU for kinetic experiments.

354

355 *In vitro IBU loading and release tests*

356 As mentioned above, the initial concentration of the IBU solution for the kinetic tests
357 was 400 mg/L; at this concentration, complete saturation of the host composite
358 material (SMNZ) is guaranteed, as suggested by the model equilibrium isotherms, and
359 greater initial concentrations of the drug do not provide further IBU uptake. Figure 6
360 highlights the rapid rate of the loading process, and > 90% of IBU was loaded onto the
361 SMNZs after 30 min, except for PBC which required ~120 min. Comparison of kinetic
362 curves reveals that the maximum amount of loaded IBU was reached for PHC and
363 PHB (26.8 and 28.1 mg/g, respectively). The latter case illustrates that a compact

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364 bilayer is not an indication of the best loading, as it can create a "crowding" of anionic
365 sites that are not all available for drug adsorption due to the larger size of IBU when
366 compared with the Br ion. The maximum loaded amounts for PBC and PCC are still
367 comparable (20.0 and 19.2 mg/g, respectively), confirming the similar behavior of
368 SMNZs modified with these two surfactants. Lastly, the similarity of the values (Table
369 4, in mEq/g) of the maximum amounts of drug loaded onto SMNZs and their
370 respective AECs (with the exception of PHB, as reported above) strongly suggests that
371 anion exchange is the primary mechanism involved in the loading of IBU on SMNZs.
372 As often reported in the literature (Bowman 2003; de Gennaro et al. 2015; Serri et al.
373 2016), the drug adsorption kinetics of SMNZs generally follow pseudo-first order
374 (PFO) (6) (Ho 2004) and pseudo-second order models (PSO) (7) (Ho 2006) . These
375 models can be written as follows:

376

377
$$Q_t = Q_0(1 - e^{-K_1 t}) \quad \text{and} \quad (6)$$

378

379
$$Q_t = \frac{K_2 Q_0^2 t}{1 + K_2 Q_0 t},$$

380 (7)

381

382 where Q_t is the amount of IBU loaded in SMNZ (mg/g) as a function of time t (min),
383 Q_0 is the drug concentration at equilibrium, and K_1 (min^{-1}) and K_2 ($\text{g/mg}^{-1} \text{min}^{-1}$) are
384 the pseudo-first and the pseudo-second order constants, respectively.

385 Non-linear regression validated both of these models (Table 4 and Fig. 6), although the
386 best fit was obtained with the pseudo-second order model, consistent with previous

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387 research on NSAIDs loaded on SMNZs (de Gennaro et al. 2015). Maximum adsorption
388 capacities were 19.3 mg/g for PCC, 20.9 mg/g for PBC, 26.4 mg/g for PHC, and 27.9
389 mg/g for PHB (calculated using the PSO model). Once again, the highest IBU
390 adsorption was achieved by PHB and PHC, whereas the amount of IBU adsorbed by
391 PCC and PBC were lower, although quite similar.

392 IBU-release kinetics, rapid and practically complete (> 95%) for all the SMNZs, were
393 fit using several mathematical models reported in the literature (Costa and Sousa Lobo
394 2001; Costa et al. 2003; Dash et al. 2010; Yadav et al. 2013), but the best fit was
395 obtained using the first-order equation (8) and the Weibull equation (9), expressed as
396 follows:

397

$$398 \quad M_t = M_0(1 - e^{-K_1 t}) \quad \text{and} \quad (8)$$

399

$$400 \quad M_t = M_0 \left[1 - e^{-\left(\frac{t-T_i}{a}\right)^b} \right], \quad (9)$$

401

402 where M_t is the amount of IBU released by SMNZ (mg/g) as a function of time t (min),
403 M_0 is the drug concentration at equilibrium (mg/g), K_1 is the first-order constant (min^{-1}),
404 T_i is the lag time (usually zero), a defines the time scale of the process (min), and b is a
405 parameter describing the shape of the dissolution profile. In particular, the curve is
406 exponential when $b = 1$, the profile becomes sigmoidal with a turning point after an
407 upward curvature if $b > 1$, and the profile is parabolic with a steeper initial slope when
408 $b < 1$.

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409 Table 5 reports the parameters of these two models fitted by non-linear regression, and
410 the dissolution profiles are shown in Figure 7, which shows that 50-60% of IBU was
411 released by all samples in the first 30 min and 85-90 % released within the first hour.
412 Nevertheless, slower drug release from the SMNZs occurred up to 5 hours, with none
413 of them achieving 100% release (Table 5).

414 Although the Weibull model is often useful for comparing dissolution profiles of
415 matrix-type drug delivery (Costa and Sousa Lobo 2001), providing good fitting results
416 (Costa et al. 2003), it represents an empirical model without any kinetic basis. The
417 Weibull model can therefore not be used to adequately describe the dissolution rate of
418 a drug or to establish effective correlations between *in vitro* and *in vivo* experiments
419 (Costa and Sousa Lobo 2001).

420 Alternatively, a first-order kinetic equation may better describe dissolution profiles of
421 IBU-loaded SMNZs, as the drug release appears to be proportional to the IBU
422 remaining in the composites (Yadav et al. 2013). In particular, much of the released
423 NSAID may be related to the amount of IBU adsorbed on the surface of SMNZs,
424 whereas minor amounts of IBU located in the hydrophobic chains of micelles would be
425 more slowly released due to diffusion processes (Pasquino et al. 2016).

426

427

IMPLICATIONS

428 One of the most important implications of this research is the possible use of low-cost
429 and largely available natural zeolites in advanced technological sectors such as
430 biomedicine. Our research has demonstrated that a phillipsite-rich rock is a good drug
431 carrier, comparable to clinoptilolite-bearing rock carriers. Our results provide added

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432 value to the southern Italian zeolite deposits, representing a by-product of the
433 building stone industry that is currently disposed in landfills. However, due to the
434 natural variability of this resource, careful characterization of production batches is
435 mandatory.

436 Experimental results provide evidence that this material, properly surface modified by
437 long-chain surfactants, is an excellent carrier of NSAIDs. Appropriate pharmaceutical
438 preparations for oral use can be developed, possibly as granulates or as dispersions in
439 order to increase the amount of the released drug. This application is supported by
440 laboratory data that have shown the prompt release of a pain-killing drug within the
441 first 30-50 min, thereby making these natural materials suitable for providing rapid
442 palliative effects to the patient. It should be remarked however that the formulation of a
443 composite system able to provide a sustained- in time drug release is required by
444 pharmacologists. The IBU-phillipsite composite material investigated in the present
445 research provided a slow release of a small amount of drug (5-10 %). Further
446 investigation is required to understand if this low amount is significant of
447 pharmacological point of view.

448 The functionalized phillipsite-rich rock may have other important applications in
449 environmental fields (Hailu et al. 2017) such as the absorption/removal of
450 pharmaceuticals from ground or surface waters, a rather growing problem which has
451 yet to find a satisfactorily solution (Larsson 2014; Płuciennik-Koropczuk 2014).

452

453

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790

Revision 2

791

792 FIGURE CAPTIONS

793

794 Figure 1 - Physico-chemical and thermal properties of IBU

795

796 Figure 2 - Physico-chemical and thermal properties of surfactants

797

798 Figure 3 - FTIR spectra of starting materials (PHI_SAV), SMNZ (PCC), IBU-loaded

799 SMNZ (PCC+IBU)

800

801 Figure 4 - Thermal properties of starting materials by TG/DSC coupled with FTIR-

802 EGA

803

804 Figure 5 - Isotherms: A-Langmuir; B-Sips; C-Toth

805

806 Figure 6 - Kinetic loading curves: A- Pseudo-first order, B- Pseudo-second order

807

808 Figure 7 - Kinetic release curves: A- First-order, B- Weibull, C- First order (%), D-

809 Weibull (%).

810

Revision 2

Table 1 – FTIR data

PHI SAV	PCC	PBC	PHC	PHB	PCC+IBU	PBC+IBU	PHC+IBU	PHB+IBU	Tentative vibrational assignments	Chemical phase
3419 w	3422 w	3436 w	3425 w	3427 w	3426 w	3434 w	3422 w	3425 w	O-H stretching	water
	2958 sh	2959 sh	2958 sh	2959 sh	2955 vw	2960 sh	2954 sh	2954 sh	C–H stretching	Surfactant/IBU
	2917 w	2923 vw	2923 vw	2923 vw	2921 w	2924 vw	2920 w	2922 w	C–H stretching	Surfactant/IBU
	2850 w	2853 vw	2852 vw	2852 vw	2851 w	2853 vw	2851 w	2852 w	C–H stretching	Surfactant/IBU
1638 w	1635 w	1639 w	1638 w	1639 w	1635 w	1638 w	1638 w	1638 w	O-H bending	water
					1572 vw	1573 sh	1580 sh	1582 sh	Asymmetric stretching in carboxylate ion	IBU
	1489 vw	1481 vw	1489 sh	1489 sh	1489 vw	1481 vw	1489 sh	1489 sh	C–H bending	Surfactant/IBU
	1469 vw	1466 vw	1470 vw	1470 vw	1466 vw	1466 vw	1469 vw	1467 vw	C–H bending	Surfactant/IBU
1456 vw									C–O asymmetric stretching	Calcite
					1382 vw	1379 vw	1376 vw	1381 vw	Symmetric stretching in carboxylate ion	IBU
1000 vs	995 vs	992 vs	995 vs	992 vs	989 vs	992 vs	991 vs	989 vs	T–O asymmetric stretching	silicates
780 w	774 w	781 w	779 w	777 w	775 w	781 w	776 w	778 w	T–O–T asymmetric stretching	silicates
719 w	717 w	723 w	720 w	719 w	719 w	724 w	720 w	719 w	T–O–T symmetric stretching	silicates
633 sh	633 sh	633 sh	632 sh	632 sh	633 sh	633 sh	633 sh	633 sh	T–O–T bending	silicates
597 w	596 w	597 w	597 w	599 w	595 w	598 w	596 w	598 w	T–O–T bending	silicates
516 vw	515 vw	519 vw	517 vw	516 vw	514 vw	517 vw	519 vw	516 vw	T–O–T bending	silicates
426 w	427 w	427 w	424 w	429 w	423 w	428 w	427 w	421 w	T–O–T bending	Silicates

Legend: w, weak; vw, very weak; sh, shoulder; s, strong; vs, very strong; T, tetrahedral cation (Si or Al).

Revision 2

Table 2 –Thermal Analysis (TG-DTG-DSC and FTIR-EGA) data

Samples	T < 200 °C			200 °C < T < 1050 °C					L.O.I (%)	R.M. (%)
	ΔW (%)	DTG (°C)	DSC ^(a) (°C)	EGA	ΔW (%)	DTG (°C)	DSC ^(b) (°C)	EGA		
PCC	6.3	157	131	H ₂ O	9.0	230-499-665	246-333-502-672 ^(a) -929	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	15.3	84.7
PCC+IBU	6.6	162	118	H ₂ O	11.0	206-488-655	223-339-498-686 ^(a) -939	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	17.5	82.5
PBC	6.1	153	164	H ₂ O	7.1	212-384-525-659	226 ^(a) -275-386-528-644 ^(a) -929	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	13.3	86.8
PBC+IBU	6.8	150	167	H ₂ O	7.7	211-384-514-646	230 ^(a) -277-388-519-646 ^(a) -932	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	14.5	85.5
PHC	6.7	144	157	H ₂ O	7.2	235-517-646	225-299-389-495-649 ^(a) -916	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	14.0	86.0
PHC+IBU	7.6	154	151	H ₂ O	8.5	216-513-648	286-380-481-645 ^(a) -943	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	16.1	83.9
PHB	6.9	146	144	H ₂ O	6.8	381-504-664	283-369-503-651 ^(a) -930	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	13.7	86.3
PHB+IBU	6.8	146	153	H ₂ O	9.1	226-385-499-665	234-296-388-476-658 ^(a) -943	CO ₂ + CH ₃ + CH ₂ + H ₂ O ^{tr} + CO ^{tr}	15.9	84.1

LEGEND:

^(a)= Endothermic

^(b)= Exothermic

L.O.I. = loss on ignition

R.M. = residual mass

ΔW = weight loss (by TG)

^(tr)= traces

Revision 1

Table 3 - Isotherm parameters

Samples	Mathematical model	Parameters				Goodness-of-fit		
		K (L/mg)	n	S _m (mg/g)	S _m (mEq/g)	R ²	AIC	BIC
PCC	Langmuir	0.032		22.0 ± 0.6	0.096	0.970	27.1	27.0
	Sips	0.026	2.91	20.4 ± 0.25	0.089	0.987	23.1	22.9
	Toth	0.014	3.85	20.4 ± 0.2	0.089	0.987	23.1	22.9
PBC	Langmuir	0.039		21.9 ± 0.5	0.096	0.981	24.2	24.1
	Sips	0.040	1.06	21.7 ± 0.4	0.095	0.981	26.1	26.0
	Toth	0.033	1.16	21.4 ± 0.4	0.094	0.982	26.0	25.9
PHC	Langmuir	0.018		26.1 ± 0.5	0.114	0.989	22.2	22.1
	Sips	0.018	1.38	24.6 ± 0.3	0.108	0.993	21.1	21.0
	Toth	0.011	1.62	24.4 ± 0.2	0.107	0.993	20.5	20.3
PHB	Langmuir	0.006		36.2 ± 1.6	0.158	0.981	29.6	29.5
	Sips	0.008	1.54	31.0 ± 0.9	0.136	0.991	26.3	26.2
	Toth	0.004	3.83	28.8 ± 0.3	0.126	0.995	21.7	21.6

Table 4 - IBU Loading kinetic runs parameters

Samples	Mathematical model	Parameters				Goodness-of-fit		
		K ₁ (min ⁻¹)	K ₂ (g·mg ⁻¹ ·min ⁻¹)	Q ₀ (mg/g)	Q ₀ (mEq/g)	R ²	AIC	BIC
PCC	Pseudo-first order	0.333		18.9 ± 1.1	0.083	0.983	27.9	28.7
	Pseudo-second order		0.041	19.3 ± 0.5	0.085	0.987	24.9	25.7
PBC	Pseudo-first order	0.073		19.2 ± 0.7	0.084	0.975	38.9	37.7
	Pseudo-second order		0.005	20.9 ± 0.5	0.091	0.979	35.2	36.0
PHC	Pseudo-first order	0.620		26.3 ± 0.4	0.115	0.996	19.3	20.1
	Pseudo-second order		0.155	26.4 ± 0.4	0.116	0.996	19.3	20.1
PHB	Pseudo-first order	0.005		27.0 ± 1.7	0.118	0.967	43.0	43.8
	Pseudo-second order		0.021	27.9 ± 0.9	0.122	0.988	31.7	32.5

Table 5 - IBU Release kinetic runs parameters

Samples	Mathematical model	Parameters					Goodness-of-fit		
		K ₁ (min ⁻¹)	a (min)	b	M ₀ (mg/g)	M ₀ (%)	R ²	AIC	BIC
PCC	First-order	0.033			18.4 ± 0.9	95.45	0.995	14.9	15.7
	Weibull		31.61	1.31	18.3 ± 0.6	94.71	0.997	9.7	10.9
PBC	First-order	0.036			19.0 ± 0.7	98.44	0.990	22.6	23.4
	Weibull		30.53	1.89	18.8 ± 0.2	97.26	0.999	-0.5	0.7
PHC	First-order	0.036			25.5 ± 1.0	96.54	0.995	21.5	22.3
	Weibull		30.08	1.51	25.3 ± 0.5	95.71	0.999	2.4	3.6
PHB	First-order	0.033			25.9 ± 1.2	92.83	0.988	31.4	32.2
	Weibull		33.19	1.68	25.6 ± 0.6	91.58	0.998	13.1	13.3

Figure 1

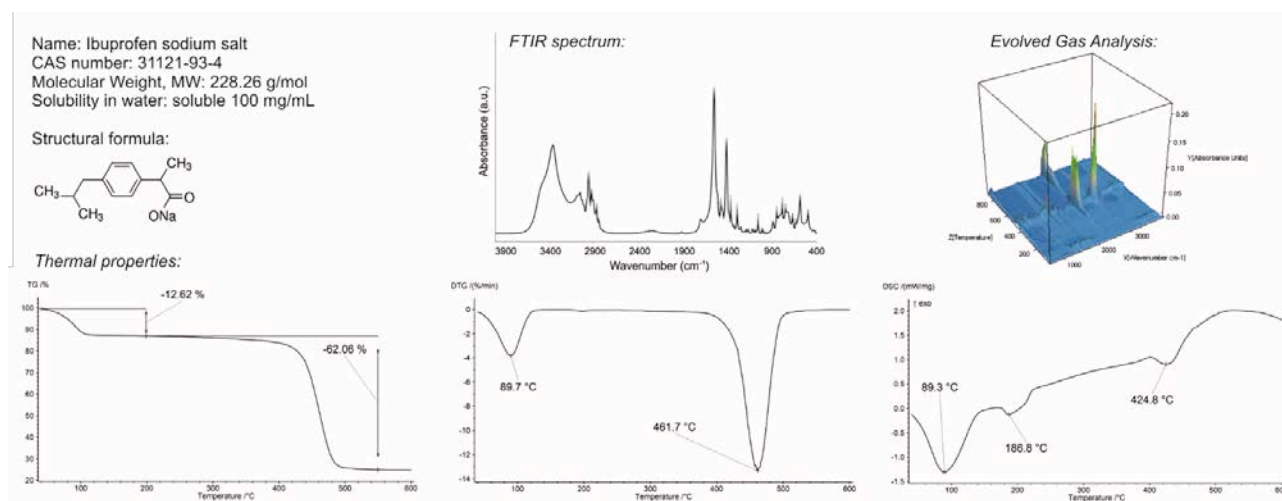
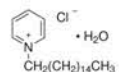


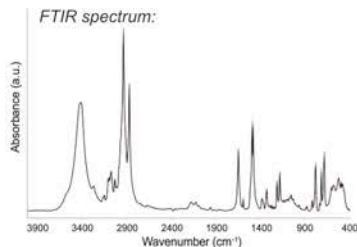
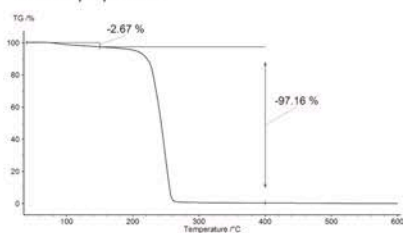
Figure 2

Name: Cetylpyridinium chloride
 CAS number: 123-03-5
 Molecular Weight, MW: 358 g/mol
 Critical Micelle Concentration, CMC: 0.9 mmol/L

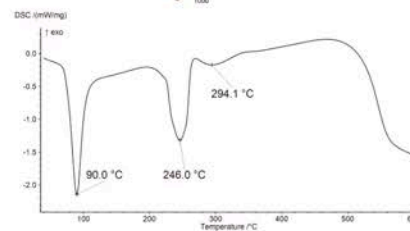
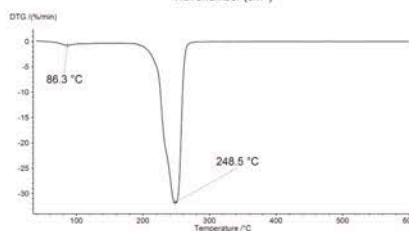
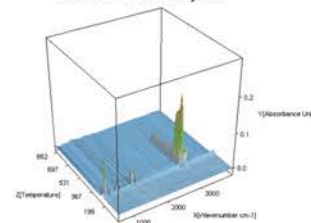
Structural formula:



Thermal properties:

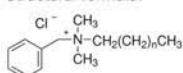


Evolved Gas Analysis:

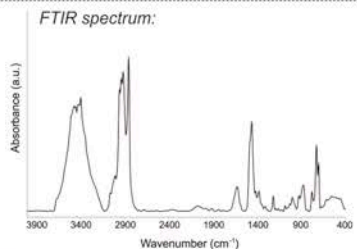
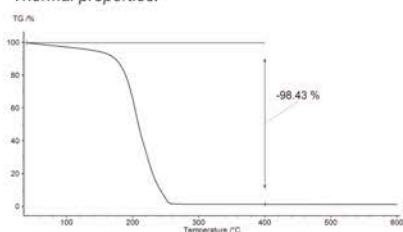


Name: Benzalkonium chloride
 CAS number: 63449-41-2
 Molecular Weight, MW: 354 g/mol
 Critical Micelle Concentration, CMC: 5 mmol/L

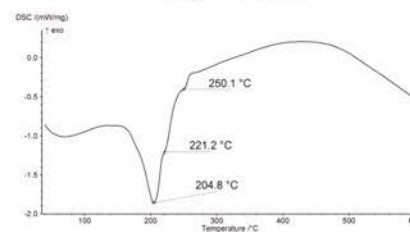
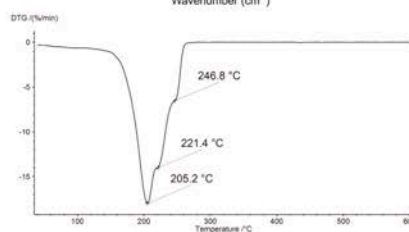
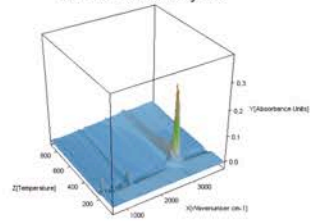
Structural formula:



Thermal properties:

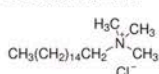


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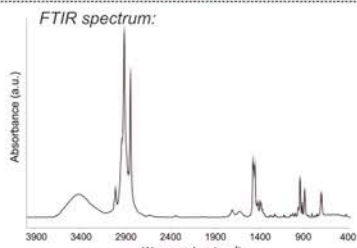
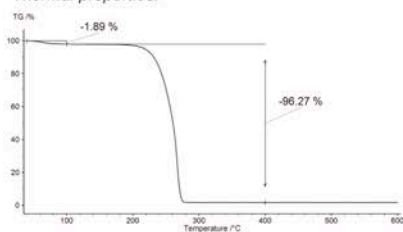


Name: Hexadecyltrimethylammonium chloride
 CAS number: 112-02-7
 Molecular Weight, MW: 320 g/mol
 Critical Micelle Concentration, CMC: 1.3 mmol/L

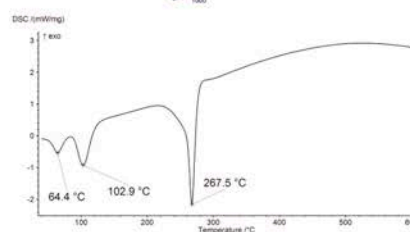
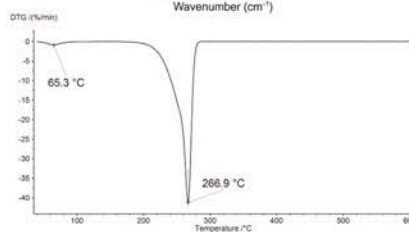
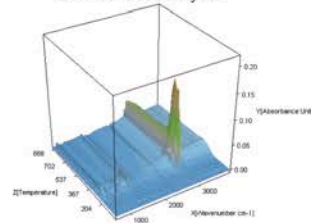
Structural formula:



Thermal properties:

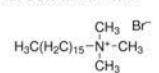


Evolved Gas Analysis:

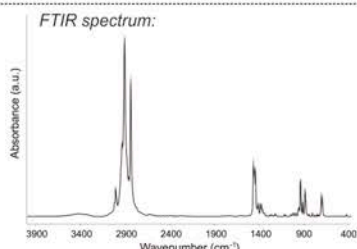
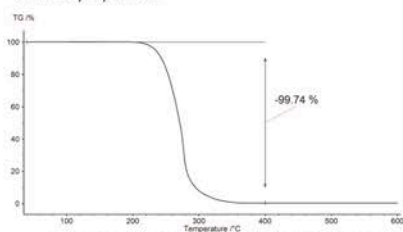


Name: Hexadecyltrimethylammonium bromide
 CAS number: 57-09-0
 Molecular Weight, MW: 364.46 g/mol
 Critical Micelle Concentration, CMC: 0.9 mmol/L

Structural formula:



Thermal properties:



Evolved Gas Analysis:

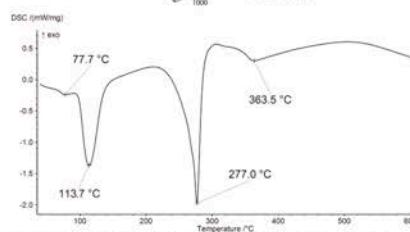
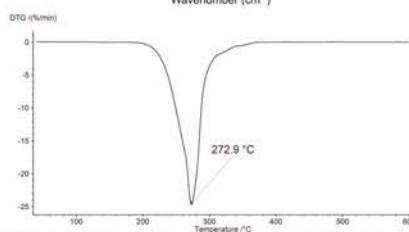
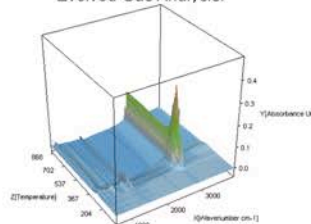


Figure 3

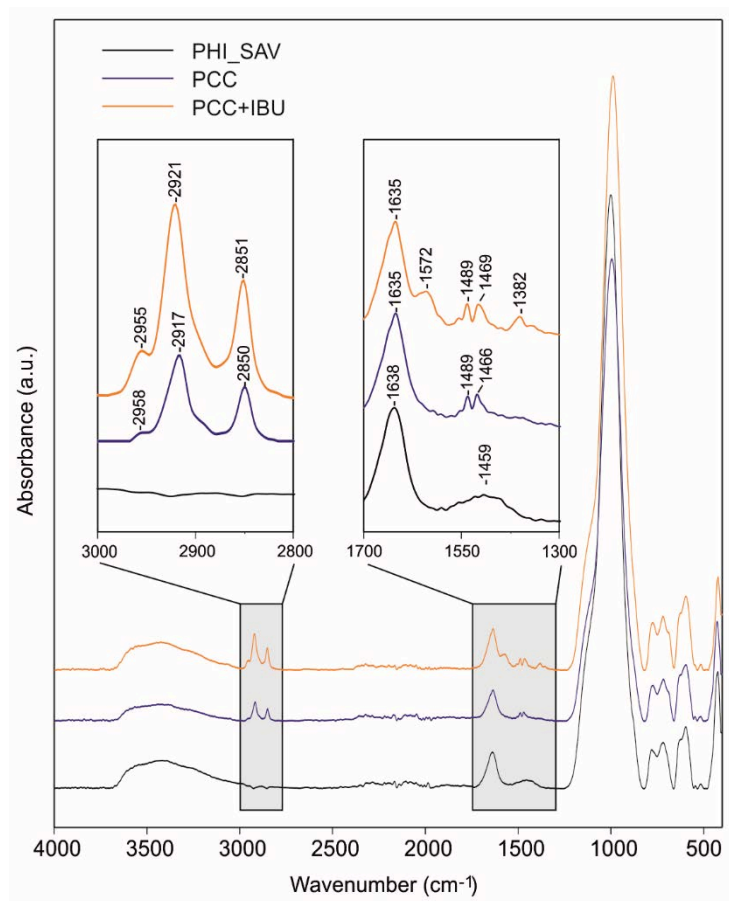


Figure 4

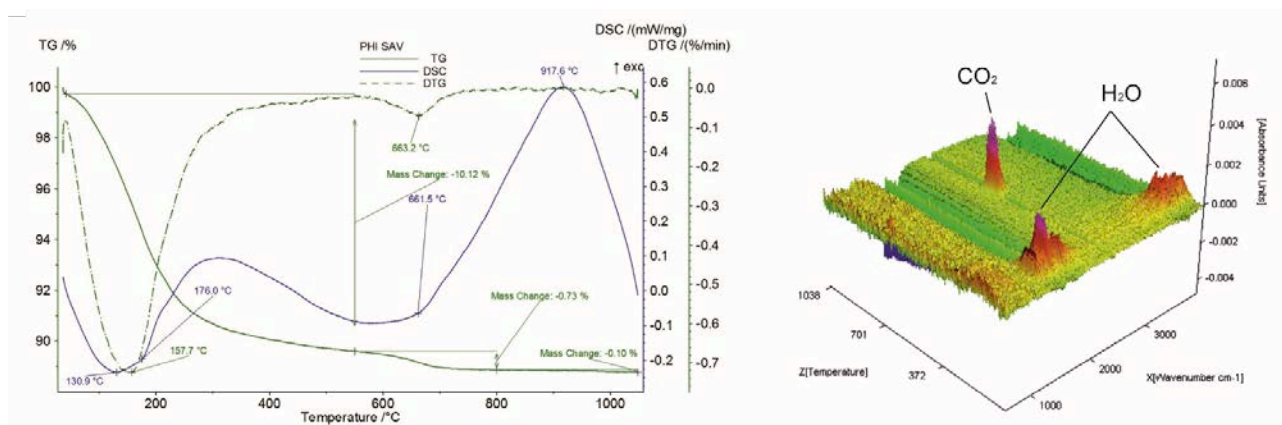


Figure 5

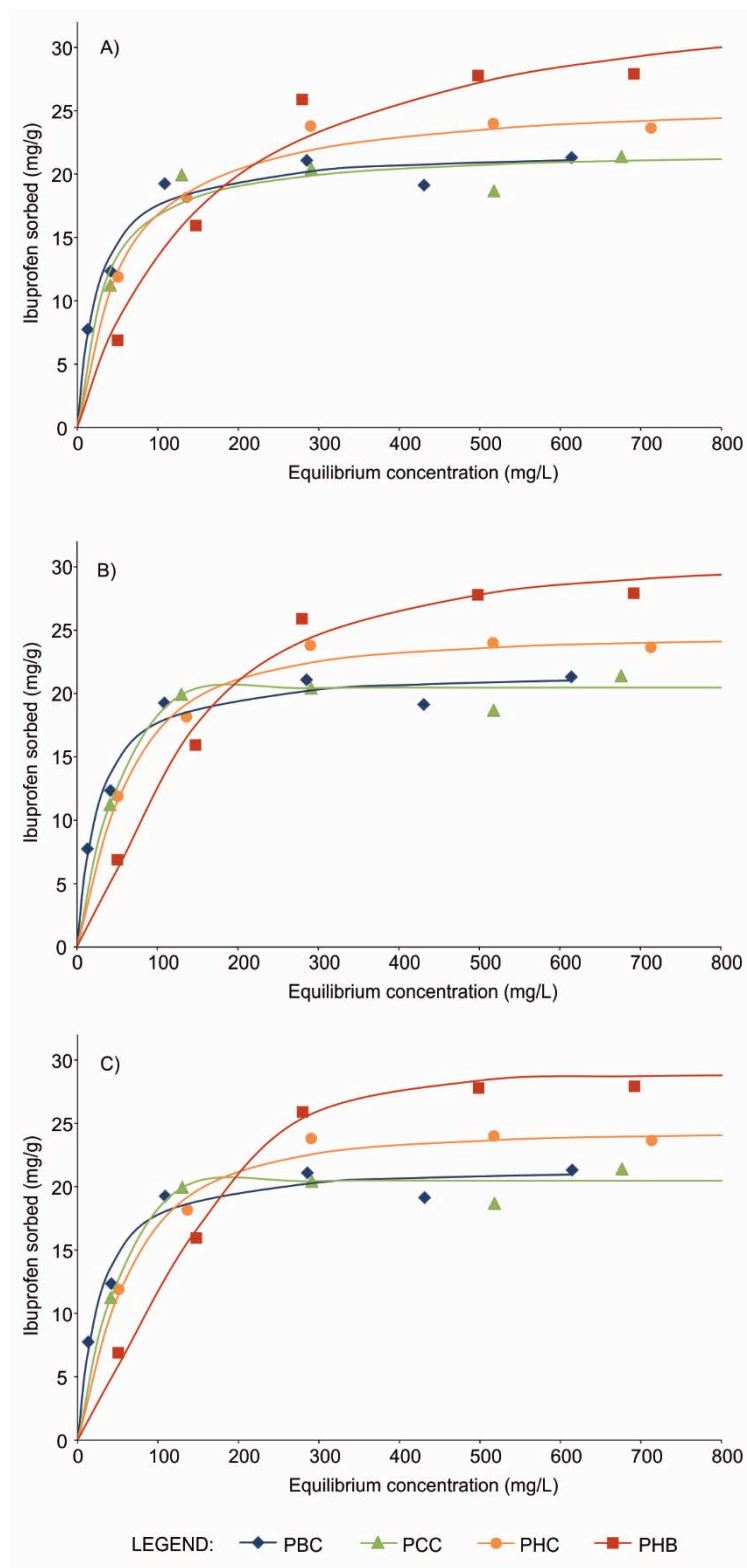


Figure 6

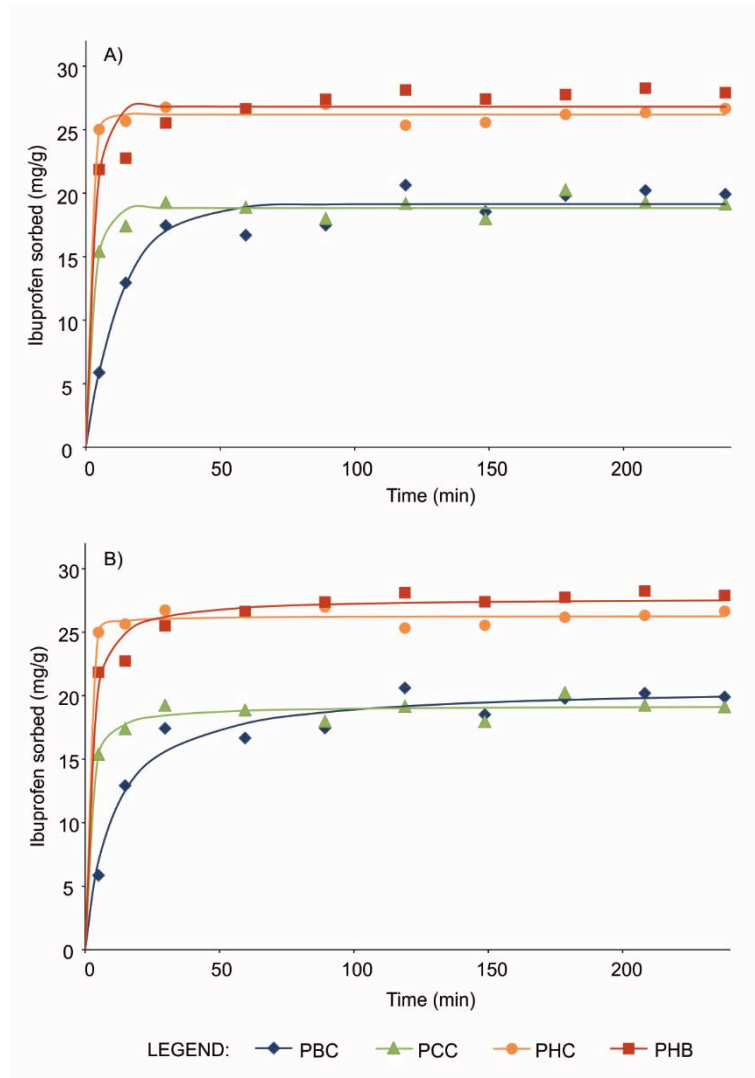


Figure 7

